



Review

# MET Inhibitors in Small Cell Lung Cancer: From the Bench to the Bedside

Max Hardy-Werbin <sup>1</sup>, Raúl del Rey-Vergara <sup>1</sup>, Miguel Alejandro Galindo-Campos <sup>1</sup>, Laura Moliner <sup>2</sup> and Edurne Arriola <sup>1,2,\*</sup>

- Cancer Research Program, IMIM (Institut Hospital del Mar d'Investigacions Mèdiques), 08003 Barcelona, Spain; mhardy@imim.es (M.H.-W.); rdelrey@imim.es (R.d.R.-V.); mgalindo@imim.es (M.A.G.-C.)
- <sup>2</sup> Medical Oncology Department, Hospital del Mar-CIBERONC, 08003 Barcelona, Spain; Imoliner@psmar.cat
- \* Correspondence: earriola@psmar.cat; Tel.: +34-932-483000

Received: 13 August 2019; Accepted: 18 September 2019; Published: 20 September 2019



**Abstract:** Small cell lung cancer (SCLC) is the most aggressive type of lung cancer. The different systemic treatment approaches attempted in the last 35 years have not improved overall survival in the advanced stage. Targeted therapies assessed in clinical trials have failed to show efficacy against SCLC. Within the potentially interesting targets, the hepatocyte growth factor (HGF)/mesenchymal-epithelial transition (MET) pathway activation is associated with worse survival and chemoresistance in SCLC. Preclinical data suggest that the inhibition of the MET pathway can revert chemoresistance and prevent tumor growth. Recently, immunotherapy has shown modest but relevant activity in SCLC. Interestingly, MET modulation seems to be involved in increasing the efficacy of standard checkpoint inhibitors. Here, we review the preclinical and clinical data of MET inhibition in SCLC, and the role of this pathway in the immune response.

Keywords: small cell lung cancer; MET; HGF; immunotherapy

### 1. Introduction

Lung cancer is the most common cancer worldwide [1]. Its incidence has increased over the years [2], and it remains the leading cause of cancer-related deaths [3]. Out of all lung cancer patients, approximately 13% are diagnosed with small cell lung cancer (SCLC) [4,5].

SCLC is the most aggressive type of lung cancer and it is highly linked to tobacco smoking [6,7]. Combination treatment with chemotherapy and radiotherapy in a limited stage has increased survival rates [8]. However, more than half of SCLC patients are diagnosed at an extensive stage, where treatments are scarce, leaving SCLC as an orphan disease with a dismal prognosis [9]. In fact, platinum-based chemotherapy has been the systemic standard treatment in an extensive disease since the mid-1980s [10,11]. Although substantial clinical responses are achieved after first-line treatment, disease progression takes place quickly and it is usually resistant to available treatments. In this scenario, SCLC presents with a 5-year overall survival (OS) of around 6% [12], dropping to 3% [4] if only the extensive disease population is considered. Thus, there is an urgent need for the development of effective therapies in SCLC.

The emergence of immunotherapy has represented the first shift in decades in the treatment of advanced SCLC, and checkpoint inhibitors have been recently approved by the Food and Drug Administration (FDA) for the treatment of this disease. Based on the results of the phase I/II trial CheckMate 032, the combination of nivolumab and ipilimumab was added as a recommendation in the National Comprehensive Cancer Network (NCCN) guidelines, and afterwards, nivolumab, as single-agent, was approved by the FDA in the third-line setting [13]. Moreover, in a recently published clinical trial (IMpower 133), the addition of atezolizumab to standard chemotherapy showed benefit

Cancers 2019, 11, 1404 2 of 14

in terms of overall survival in chemo-naive SCLC patients [14]. Nevertheless, only a small subset of unselected patients seems to benefit from immunotherapy. Despite several biomarkers that have been evaluated as predictive markers showing benefit from immunotherapy in SCLC [15,16], these have not yet been validated.

SCLC is characterized as having one of the highest rates of mutational tumor burden [17]. However, despite the multiple genomic aberrations found in SCLC [18], these are rarely druggable. Although many attempts have been made, targeted therapies have not shown benefits regarding SCLC treatment [19,20]. One of the potentially interesting targets, linked to mechanisms of chemoresistance in SCLC, is the activation of the hepatocyte growth factor (HGF)/mesenchymal-epithelial transition (MET) receptor tyrosine kinase (RTK) signaling pathway via the induction of the epithelial to mesenchymal transition (EMT). In preclinical models it has been shown that this pathway plays a key role in the aggressiveness of SCLC [21] and that its inhibition leads to the reversion of chemoresistance and tumor response [22]. This review discusses the role of the HGF/MET pathway in SCLC and the potential therapeutic value of MET inhibition in this deadly disease.

# 2. Description of the MET Molecule and the Downstream Pathway

The human proto-oncogene MET was originally described in 1984 [23]. After treating a human osteosarcoma cell line with a chemical carcinogen (N-methyl-N0-nitro-N-nitrosoguanidine), a fusion protein with the translocated promoter region (TPR) and the MET domain was obtained. The TPR region of the protein induced a constitutive dimerization, which led to an activation of the MET kinase domain. MET is located in chromosome 7q21-q31 [24], and codifies to a high-affinity transmembrane RTK [25]. The MET receptor is a disulphide linked glycoprotein heterodimeric RTK. The structure of the molecule consists of an extracellular  $\alpha$ -chain linked to transmembrane-spanning  $\beta$ -chain. The  $\alpha$ -chain and the first fragments of the  $\beta$ -chain are responsible for the binding to its ligand, the HGF, while the intracellular region of the  $\beta$ -chain is responsible of the downstream signaling [26].

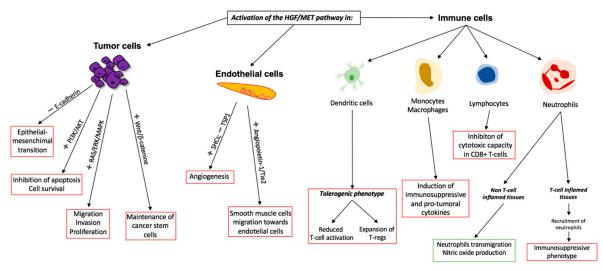
By the time the MET receptor was characterized, its ligand had not yet been identified [27]. Later, HGF was described by two independent groups as a motility and a mitogenic factor [28,29], and then defined as the MET ligand [30]; nowadays, it remains as the sole identified ligand of the MET receptor.

After binding to HGF, the MET receptor suffers a dimerization [31], which causes a phosphorylation in the tyrosine residues of the cytoplasmic domain, mainly at phospho-epitopes pY1230, pY1234, and pY1235 [32,33]. Afterwards, phosphorylation in pY1349 promotes the recruitment of several substrates, such as growth factor receptor-bound protein 2 (Grb2), Gab1 (Grb2-associated-binding protein 1), Src and phospholipase C [34], that are responsible for the following downstream signaling, essentially through the rat sarcoma (RAS)-mitogen-activated protein kinase (MAPK) and phosphoinositide 3-kinase (PI3K)-protein kinase B (AKT) pathways [26,32,35,36].

Under physiological conditions, the HGF/MET pathway mediates several cellular responses, such as cell migration [36] and tissue regeneration [37,38]. In this context, MET activation is basically controlled in two different ways: 1) internalization by endocytosis into multivesicular bodies, followed by ubiquitination and degradation [39,40]; and 2) fading of signaling due to the action of specific phosphatases [41].

However, aberrant activation of the MET pathway leads to increased cell motility and cell cycle progression and many other protumorigenic signaling (Figure 1), and has been proven to be involved in many types of cancer, as discussed below.

Cancers 2019, 11, 1404 3 of 14



**Figure 1.** Role of the activation of the HGF/MET axis in tumor cells, endothelial cells, and immune cells. Red boxes define pro-tumorogenic functions. Green boxes define anti-tumorogenic functions. PI3K, phosphoinositide 3-kinase; ERK, extracellular signal-regulated kinase; MAPK, mitogen-activated protein kinase; SHCs—SRC homology 2 domain-containing proteins; TSP1—thrombospondin 1; Tie2—angiopoietin receptor 2.

## 3. Role of the HGF/MET Axis in Cancer

During the last three decades, the dysregulation of the HGF/MET pathway has been identified as a common characteristic in different types of human cancer. In the early 1990s, Rong et al. proved the tumorigenic feature of the oncogene *MET* and the metastatic potential of the overexpression of HGF in murine and human cancer cell lines [42,43]. This fact was confirmed by a loss of aggressiveness when the HGF/MET axis was downregulated in human tumors [44]. Later on, many studies showed that HGF and MET were often expressed in a large variety of cancers [45–50].

The MET downstream signaling can be aberrantly enhanced by gene mutation or amplification, by protein overexpression, or in a ligand-dependent fashion. In 1997, Schmidt et al. [51] reported, for the first time, activating mutations of MET in renal cancer. Further studies found other MET mutations in many cancer types, including SCLC [52,53], supporting the implication of this receptor in human cancer [54–58]. Some of these mutations were later identified as non-pathogenic, thus more research is required in this field [21].

Among the acquired functions due to the aberrant activation of MET, the epithelial-mesenchymal transition (EMT) is of pronounced significance. EMT is a phenomenon whereby cancer cells suffer a loss of epithelial features and acquire a mesenchymal phenotype, conferring the capability to go through the boundaries of the tumor and metastasize [59]. The driver characteristic of this process is the suppression of E-cadherin, which is mainly induced by the expression of the transcriptional factor Snail. It has been reported that HGF is an effective inductor of EMT [60,61]. Our group has previously demonstrated that activation of the HGF/MET pathway induced an EMT phenotype in a SCLC preclinical model [21]. Moreover, MET inhibition was able to revert the mesenchymal features on these cells.

The HGF/MET axis also plays a crucial role in resistance to targeted drugs. It has been show that HGF can induce tyrosine-kinase inhibitors resistance in lung adenocarcinoma harboring epidermal growth factor receptor (EGFR) mutations, presumably through cross-talk activation of the PI3K/AKT pathway [62,63].

In addition to its role in tumor cells, MET activation can also be observed in endothelial cells, promoting angiogenesis [64,65], thus contributing to tumor progression. Through the regulation of diverse intermediates, such as SHCs (SRC homology 2 domain-containing proteins) and TSP1 (thrombospondin), MET favors the induction of endothelial cell growth [66] (Figure 1).

Cancers 2019, 11, 1404 4 of 14

Activation of the HGF/MET axis has not only been shown to be a marker of aggressiveness in preclinical research. Several groups reported a decreased survival rate in cancer patients with an aberrant tumor expression of either MET or its ligand HGF, including SCLC patients [67–70].

#### 4. MET in SCLC

## 4.1. Emergence of MET in the SCLC Biomarker Spectrum

The first study to show the expression of MET in SCLC was conducted by Rygaard et al. in 1993 [71]. They examined a panel of 25 SCLC cell lines and xenografts, demonstrating the presence of MET mRNA transcripts and protein expression, but failing to show that these same tumor cells co-expressed its ligand, HGF.

The role of the MET/HGF axis in SCLC was later elucidated via stimulation of SCLC cell lines with HGF. Maulik et al. found an increased tyrosine phosphorylation of MET, and an increase in cell motility (observed by the formation of filopodia) in response to HGF [72]. They also showed that the heat shock protein 90 (Hsp90) inhibitor geldanamycin, a known inhibitor of the HGF/MET pathway [73], increased apoptosis, and decreased motility and MET expression in SCLC cell lines. Further research supported this concept by showing that the C-terminus of the Hsp70-interacting protein (CHIP), which also targets Hsp90, is essential for MET regulation by blocking cell survival pathways in SCLC cell-lines [74]. Similar effects were seen when inhibiting PI3K, a downstream factor of MET, in MET-dependent SCLC cell lines [75].

#### 4.2. Small MET Inhibitor Molecules

In light of the relevance of the HGF/MET axis activation in SCLC, several novel MET inhibitors were tested. SU11274 is a small molecule inhibitor of MET and it has been proved to be 50 times more selective for MET relatively to other kinases [76]. SU11274 selectively inhibited MET tyrosine kinase activity in a cell line transformed by the oncogenic TPR-MET protein (a feature also present in SCLC), inducing cell cycle arrest and apoptosis [77]. This inhibition was specifically replicated in non-small-cell lung cancer NSCLC and SCLC cell lines and supported by obtaining the same results after the inhibition of MET via knock-down with a small interference RNA (siRNA) [33,78].

MET targeting drugs were later tested in vivo. PHA665752 is a specific, small-molecule MET ATP-competitive inhibitor that also synergizes with the downstream specific mTOR inhibitor rapamycin [79]. SCLC xenograft mouse models treated with PHA665752 showed high rates of tumor response [80]. Of note, these mice were injected with the drug intratumorally. Further in vitro assays with SCLC cell lines were able to support the concept regarding how PHA665752 counteracts the stimulation of HGF in *MET* mutant SCLC [22].

# 4.3. Combination of MET Inhibitors with Chemotherapy

Once specific MET inhibitors were shown to be active in SCLC, further new strategies were evaluated. Prof. Salgia's group [81] aimed to look for combination strategies with chemotherapeutic agents and MET inhibitors. They found that extensive SCLC patients overexpressed nuclear topoisomerase-1, and that this fact was correlated with the increased expression of MET. Thus, they tested the MET inhibitor SU11274 in vitro in combination with 7-Ethyl-10-hydroxy-camptothecin (SN-38), the active metabolite of irinotecan [82], a topoisomerase inhibitor used in refractory disease in SCLC [83]. Both compounds combined synergized together, achieving a greater decrease on cell viability in the SCLC cell line H69 than when used alone.

Shortly after, a multikinase inhibitor was tested by our group [21] in combination with the topoisomerase-2 inhibitor etoposide. PF-2341066 (crizotinib), a small molecule inhibitor, showed activity against the anaplastic lymphoma RTK (ALK) and MET [84]. In order to establish the role of the MET/HGF axis activation in chemoresistance, we were able to resensitize chemoresistant SCLC cell lines, both in vitro and in vivo after treatment with PF-2341066. Moreover, a significant decrease in

Cancers 2019, 11, 1404 5 of 14

tumor growth was observed on H69M (mesenchymal cells derived from H69 after HGF exposure) xenografted mice after treatment with PF-2341066 combined with etoposide [21]. These findings were subsequently validated by different groups. Ozasa et al. [85] reported that treating SCLC chemoresistant cell lines with the MET inhibitor SU11274 combined with irinotecan reverted the resistance to cytotoxic treatment. In a preclinical scenario from an orthotopic SCLC model with extensive disease, Sakamoto et al. showed that treatment with the Met inhibitor PHA665752 inhibited motility and the invasion of SCLC cells harboring high HGF expression [86]. In this case, combination with chemotherapy was not assessed. In a more recent work, Taniguchi et al. [87] also showed similar results, both in vitro and in vivo, with E7050 (golvatinib), a dual kinase inhibitor of MET and vascular endothelial growth factor receptor-2 (VEGFR-2), which is currently under clinical development [88]. Growth of SCLC cell lines expressing HGF and phosphorylated-MET (confirmed using Western blot and Enzyme-linked immunosorbent assay) was constrained after in vitro treatment with golvatinib. Moreover, this MET inhibitor was able to avoid systemic SCLC metastases in vivo. In this study, the proof of concept was supported by obtaining similar results with a knockdown of HGF and MET. Of note, the same results were observed using crizotinib. Combinations with chemotherapeutic agents was not evaluated. These data suggest a role for MET inhibition in SCLC (Table 1).

Anti-MET Drug	Regimen	Reference	Test Setting
NSC122750 (geldanamycin)	Monotherapy	Maulik et al. (2002) [64]	In vitro
SU11274	Monotherapy	Sattler et al. (2003) [69] Ma et al. (2007) [31] Ma et al. (2005) [70]	In vitro In vitro In vitro
SU11274	Combination with irinotecan	Rolle et al. (2013) [74] Osaza et al. (2014) [78]	In vitro In vitro
PHA665752	Monotherapy	Puri et al. (2007) [72] Arriola et al. (2011) [20] Sakamoto et al. (2015) [79]	In vivo In vitro In vitro
PF2341066 (criozotinib)	Combination with etoposide	Cañadas et al. (2014) [19] Taniguchi et al. (2017) [80]	In vitro/in vivo In vitro/in vivo
E7050 (golvatinib)	Monotherapy	Taniguchi et al. (2017) [80]	In vitro/in vivo

**Table 1.** Preclinical testing of anti-MET targeted therapy in SCLC.

#### 4.4. Clinical Trials

In spite of all the preclinical research of the efficacy of anti-MET therapy in SCLC developed in the last two decades, few clinical trials have been conducted, of which none delivered positive results.

Tivantinib (ARQ-197) is a small-molecule inhibitor of MET [89]. However, its main mechanism of action may involve a tubulin depolymerization [90]. It has been tested in a phase I clinical trial [91], in combination with topotecan, in patients with advanced solid tumors, including three patients with SCLC. An expansion cohort consisting of advanced stage SCLC was planned, but development was halted due to poor tolerability of the combination. Moreover, no responses were seen. During the course of this trial, a concomitant phase II clinical trial with tivantinib as a maintenance treatment in advanced stage SCLC was initiated, but suffered an early termination due to the preliminary safety results [92].

Efficacy of rilotumumab (anti-HGF) or ganitumab (anti-insulin-like growth factor 1 receptor) in combination with platinum-etoposide was assessed in first-line treatment in advanced stage SCLC. A phase Ib/II clinical trial [93] failed to show improved outcomes with the addition of an anti-HGF monoclonal antibody to chemotherapy. This was a two-part study: the first part was a dose escalation phase for both inhibitors; and the second part was a randomized placebo-controlled phase using the doses identified in the first part where patients were assigned to receive rilotumumab, ganitumab, or placebo, plus chemotherapy. The overall survival in the anti-HGF group was higher than the

Cancers 2019, 11, 1404 6 of 14

placebo group (12.2 vs 10.8 months), although this difference was not statistically significant. Objective response rate (ORR) was 59% in the placebo group and 68% in the rilotumumab group.

Finally, a phase II clinical trial [94] evaluating the efficacy of amuvatinib, a multi-targeted tyrosine kinase inhibitor, in combination with platinum-etoposide in relapsed SCLC patients was conducted. Although the study was designed mainly because of the activity of the inhibitor in c-KIT mutant forms, it also has activity against MET. However, the trial failed to meet its primary objective, which discouraged further development of the drug in this scenario.

# 4.5. Prevalence of MET Alterations in SCLC

The data available to this day regarding the importance of aberrant activation of the HGF/MET axis in SCLC supports the evaluation of the inhibition of this pathway as a therapeutic strategy. Initially, Ma et al. reported a prevalence of 12.5% in a cohort of 32 SCLC tumor samples [52]: only one mutation (E168D) was shown to elicit a gain of functions. Later on, Pfeifer et al. did an integrative genome analyses in 99 SCLC samples and did not find any MET pathogenic mutation [95]. More recently, two studies reported a MET mutation prevalence of 6.5% and 4.4% in 46 and 113 SCLC tumor specimens, respectively [96,97]. Globally, the prevalence of MET mutations in SCLC varies among the different studies, and most of the mutations were found to be non-pathogenic. Other mechanisms, such as increased HGF secretion, might identify a subset of patients eligible for effective MET inhibition [21]. Interestingly, a recent publication highlights the relevance of selective recognition and imaging of two-chain active HGF using a macrocyclic peptide that might serve as a biomarker for MET-activated tumors targetable with MET inhibitors [98]. The complexity of the MET receptor ligand's dependent and independent activation will require comprehensive biomarker studies when testing MET inhibitors in this disease.

#### 5. Rationale for MET Inhibition in Combination with Immunotherapy

There is increasing evidence of the role of the HGF/MET signaling in immune responses, and its potential effects in the immune compartment of the tumor microenvironment (Figure 1).

As mentioned before, MET is highly expressed in many cancer types, acting as an epitope. As such, it can be recognized by cytotoxic CD8+ T cells, eliciting an activation toward tumor cells expressing MET [99].

On the other hand, MET is not only expressed by cancer cells, but also in stromal and immune cells [100,101]. It plays a crucial role involving dendritic cells (DCs). MET activation through HGF stimulation in DCs may lead to a tolerogenic phenotype with the consequent proliferation of regulatory CD4+ T cells and a decrease of cytotoxic CD8+ T cells, resulting in immunosuppression [102]. In the T cell compartment, it has been recently reported that a fraction of CD8+ T cells express MET, and that upon stimulation with HGF, their cytotoxic capacity is restrained [103].

MET is also expressed in neutrophils. Finisguerra et al. [104] showed that MET deletion in neutrophils in a murine model enhances tumor growth and metastasis. This suggests that MET constitutes a relevant requirement for neutrophils chemoattraction and cytotoxicity in order to trigger an anti-tumor local immune response, mainly through an HGF/MET-dependent cytotoxic nitric oxide release. These findings might be relevant when treating SCLC with MET inhibitors regardless of MET status, in which the neutrophil mediated cytotoxicity may play a significant role.

Even though these findings call for caution, the scenario changes when immunotherapy (anti-programmed death(PD)-1 agents) is added into the treatment regime. In a tumoral context, interleukin (IL)-17 from  $\gamma\delta$  T cells induce neutrophils' ability to suppress cytotoxic CD8+ T cells [105]. Glodde et al. [106] showed in immune-competent mice treated with anti-PD-1, that neutrophils recruited to the tumor acquired an immunosupressive phenotype upon exposure to interferon (IFN) $\gamma$ , which was mainly released by activated T cells in the tumor microenvironment. The addition of a MET inhibitor to immunotherapy blocked the reactive migration of neutrophils from the bone marrow to the tumor, achieving better responses to anti-PD-1 agents. These results were replicated in mice

Cancers 2019, 11, 1404 7 of 14

with tumors harboring MET aberrant signaling, as well as in mice with wild-type MET, reinforcing the potential efficacy of a combination strategy of immunotherapy and anti-MET inhibitors, regardless of the tumor MET status.

There is recent evidence suggesting that patients with MET exon 14 mutant lung cancer respond worse to immunotherapy [107,108], further supporting exploring the combination. However, preclinical and clinical data evaluating the combination of immunotherapy and MET inhibitors in SCLC are still lacking.

#### 6. Conclusions

SCLC is a highly lethal malignancy with a dismal prognosis and little improvements have been achieved to date. In the last few years, the advent of immunotherapy has allowed us to witness the emergence of a small percentage of patients presenting a longer survival than expected. However, targeted therapies failed to show any progress and none of them are available for the treatment of patients with SCLC.

There are currently around 120 clinical trials in active recruitment worldwide involving targeted therapies and immunotherapy in SCLC, including those basket trials for solid tumors, allowing for the inclusion of SCLC patients. Of those, none of them includes drugs targeting MET to our knowledge.

In the past, only two trials attempted to block MET with a selective target SCLC population, and both were negative. Diverse explanations can be hypothesized for this failure. First, the anti-MET agents used had scarce preclinical data on SCLC. Rilotumumab (AMG-102) is a monoclonal antibody that prevents the HGF binding to the MET receptor; to the best of our knowledge there are no reports on preclinical data of rilotumumab in SCLC. Furthermore, given the consecutive negative results in several clinical trials and its increased toxicity, its mechanism of action was further studied. Greenall et al. [109] reported that rilotumumab does not prevent HGF from directly binding to HGF, which maybe explains the negative trial results. Tivantinib (ARQ-197), a selective non-ATP-competitive inhibitor of MET, also failed to demonstrate efficacy in SCLC, and it showed intolerable toxicity when combined with chemotherapy. Preclinical data were lacking evaluating activity of this molecule in MET-dependent SCLC models [110]. Furthermore, it has been reported that its efficacy is independent of MET inhibition due to its primary mechanism of action via tubulin depolymerization [90], which highlights the relevance of thorough preclinical and mechanistic studies before initiating a clinical trial. Notably, MET status in patients recruited in these clinical trials had not been assessed, leading one to suspect that the population was not properly selected and therefore may mask the benefit for MET driven tumors.

Immunotherapy has shifted how cancer patients are treated nowadays, and SCLC patients are no exception. A combination of immunotherapy and MET-inhibitors seems a potentially promising strategy to be assessed regarding SCLC, irrespective of patients harboring MET overexpression/mutation. Adequate selection of patients for clinical trial success will require biomarker studies including MET activation, HGF expression, immune infiltrate evaluation, and others.

In conclusion, we believe there is rationale for the evaluation of MET inhibitors with immunotherapy in SCLC. It is now time to design proper clinical trials and take this knowledge from the bench to the bedside.

Author Contributions: M.H.-W., R.d.R.-V., M.A.G.-C., L.M., and E.A. conceived and revised the paper.

Funding: This research received no external funding.

Conflicts of Interest: Edurne Arriola has received: honoraria for consultancy and lectures from Bristol-Myers Squibb, Roche, Merck Sharp & Dohme, Pfizer, Lilly, Astrazeneca, and Boeringer; research funding from Roche and Pfizer (outside the current work); institutional funding from Bristol-Myers Squibb, Pfizer and Roche (outside the current work); and travel and educational expenses from Bristol-Myers Squibb, Merck Sharp & Dohme, Roche, and Lilly. The remaining authors report no conflict of interest.

Cancers 2019, 11, 1404 8 of 14

#### References

1. Ferlay, J.; Soerjomataram, I.; Dikshit, R.; Eser, S.; Mathers, C.; Rebelo, M.; Parkin, D.M.; Forman, D.; Bray, F. Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int. J. Cancer* 2015, 136, E359–E386. [CrossRef] [PubMed]

- 2. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 2018, 68, 394–424. [CrossRef] [PubMed]
- 3. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2019. *CA Cancer J. Clin.* **2019**, 69, 7–34. [CrossRef] [PubMed]
- 4. Howlader, N.; Noone, A.M.; Krapcho, M.; Miller, D.; Bishop, K.; Kosary, C.L.; Yu, M.; Ruhl, J.; Tatalovich, Z.; Mariotto, A.; et al. *Cancer Statistics Review*, 1975–2014; National Cancer Institute: Bethesda, MD, USA, 2017.
- Govindan, R.; Page, N.; Morgensztern, D.; Read, W.; Tierney, R.; Vlahiotis, A.; Spitznagel, E.L.; Piccirillo, J. Changing epidemiology of small-cell lung cancer in the United States over the last 30 years: Analysis of the surveillance, epidemiologic, and end results database. *J. Clin. Oncol.* 2006, 24, 4539–4544. [CrossRef] [PubMed]
- 6. Khuder, S.A. Effect of cigarette smoking on major histological types of lung cancer: A meta-analysis. *Lung Cancer* **2000**, *31*, 139–148. [CrossRef]
- 7. Ou, S.H.I.; Ziogas, A.; Zell, J.A. Prognostic factors for survival in extensive stage small cell lung cancer (ED-SCLC): The importance of smoking history, socioeconomic and marital statuses, and ethnicity. *J. Thorac. Oncol.* 2009, 4, 37–43. [CrossRef] [PubMed]
- 8. Puglisi, M.; Dolly, S.; Faria, A.; Myerson, J.S.; Popat, S.; O'Brien, M.E.R. Treatment options for small cell lung cancer—Do we have more choice? *Br. J. Cancer* **2010**, *102*, 629–638. [CrossRef]
- 9. Nicholson, A.G.; Chansky, K.; Crowley, J.; Beyruti, R.; Kubota, K.; Turrisi, A.; Eberhardt, W.E.E.; Van Meerbeeck, J.; Rami-Porta, R. The international association for the study of lung cancer lung cancer staging project: Proposals for the revision of the clinical and pathologic staging of small cell lung cancer in the forthcoming eighth edition of the tnm classification for lung cancer. *J. Thorac. Oncol.* **2016**, *11*, 300–311. [CrossRef]
- 10. Oze, I.; Hotta, K.; Kiura, K.; Ochi, N.; Takigawa, N.; Fujiwara, Y.; Tabata, M.; Tanimoto, M. Twenty-seven years of phase III trials for patients with extensive disease small-cell lung cancer: Disappointing results. *PLoS ONE* **2009**, *4*, e7835. [CrossRef]
- 11. Sabari, J.K.; Lok, B.H.; Laird, J.H.; Poirier, J.T.; Rudin, C.M. Unravelling the biology of SCLC: Implications for therapy. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 549. [CrossRef]
- 12. Wang, S.; Tang, J.; Sun, T.; Zheng, X.; Li, J.; Sun, H.; Zhou, X.; Zhou, C.; Zhang, H.; Cheng, Z.; et al. Survival changes in patients with small cell lung cancer and disparities between different sexes, socioeconomic statuses and ages. *Sci. Rep.* **2017**, *7*, 1339. [CrossRef] [PubMed]
- 13. Antonia, S.J.; López-Martin, J.A.; Bendell, J.; Ott, P.A.; Taylor, M.; Eder, J.P.; Jäger, D.; Pietanza, M.C.; Le, D.T.; de Braud, F.; et al. Nivolumab alone and nivolumab plus ipilimumab in recurrent small-cell lung cancer (CheckMate 032): A multicentre, open-label, phase 1/2 trial. *Lancet Oncol.* 2016, 17, 883–895. [CrossRef]
- 14. Horn, L.; Mansfield, A.S.; Szczęsna, A.; Havel, L.; Krzakowski, M.; Hochmair, M.J.; Huemer, F.; Losonczy, G.; Johnson, M.L.; Nishio, M.; et al. First-Line Atezolizumab plus Chemotherapy in Extensive-Stage Small-Cell Lung Cancer. *N. Engl. J. Med.* **2018**, *379*, 2220–2229. [CrossRef] [PubMed]
- 15. Hardy-Werbin, M.; Rocha, P.; Arpi, O.; Taus, Á.; Nonell, L.; Durán, X.; Villanueva, X.; Joseph-Pietras, D.; Nolan, L.; Danson, S.; et al. Serum cytokine levels as predictive biomarkers of benefit from ipilimumab in small cell lung cancer. *Oncoimmunology* **2019**, *8*, e1593810. [CrossRef] [PubMed]
- 16. Hardy-Werbin, M.; Arpí, O.; Taus, A.; Rocha, P.; Joseph-Pietras, D.; Nolan, L.; Danson, S.; Griffiths, R.; Lopez-Botet, M.; Rovira, A.; et al. Assessment of neuronal autoantibodies in patients with small cell lung cancer treated with chemotherapy with or without ipilimumab. *Oncoimmunology* **2018**, 7, e1395125. [CrossRef] [PubMed]
- 17. Alexandrov, L.B.; Nik-Zainal, S.; Wedge, D.C.; Aparicio, S.A.J.R.; Behjati, S.; Biankin, A.V.; Bignell, G.R.; Bolli, N.; Borg, A.; Børresen-Dale, A.L.; et al. Signatures of mutational processes in human cancer. *Nature* **2013**, *500*, 415–421. [CrossRef] [PubMed]

Cancers 2019, 11, 1404 9 of 14

18. George, J.; Lim, J.S.; Jang, S.J.; Cun, Y.; Ozretia, L.; Kong, G.; Leenders, F.; Lu, X.; Fernández-Cuesta, L.; Bosco, G.; et al. Comprehensive genomic profiles of small cell lung cancer. *Nature* **2015**, *524*, 47–53. [CrossRef]

- 19. Ready, N.E.; Pang, H.H.; Gu, L.; Otterson, G.A.; Thomas, S.P.; Miller, A.A.; Baggstrom, M.; Masters, G.A.; Graziano, S.L.; Crawford, J.; et al. Chemotherapy with or without Maintenance Sunitinib for Untreated Extensive-Stage Small-Cell Lung Cancer: A Randomized, Double-Blind, Placebo-Controlled Phase II Study—CALGB 30504 (Alliance). *J. Clin. Oncol.* 2015, 33, 1660–1665. [CrossRef]
- 20. Arnold, A.M.; Seymour, L.; Smylie, M.; Ding, K.; Ung, Y.; Findlay, B.; Lee, C.W.; Djurfeldt, M.; Whitehead, M.; Ellis, P.; et al. Phase II study of vandetanib or placebo in small-cell lung cancer patients after complete or partial response to induction chemotherapy with or without radiation therapy: National Cancer Institute of Canada Clinical Trials Group Study BR.20. J. Clin. Oncol. 2007, 25, 4278–4284. [CrossRef]
- 21. Cañadas, I.; Rojo, F.; Taus, Á.; Arpí, O.; Arumí-Uría, M.; Pijuan, L.; Menéndez, S.; Zazo, S.; Dómine, M.; Salido, M.; et al. Targeting epithelial-to-mesenchymal transition with Met inhibitors reverts chemoresistance in small cell lung cancer. *Clin. Cancer Res.* **2014**, *20*, 938–950. [CrossRef]
- 22. Arriola, E.; Cañadas, I.; Arumí-Uría, M.; Dómine, M.; Lopez-Vilario, J.A.; Arpí, O.; Salido, M.; Menéndez, S.; Grande, E.; Hirsch, F.R.; et al. MET phosphorylation predicts poor outcome in small cell lung carcinoma and its inhibition blocks HGF-induced effects in MET mutant cell lines. *Br. J. Cancer* 2011, 105, 814–823. [CrossRef] [PubMed]
- 23. Cooper, C.S.; Park, M.; Blair, D.G.; Tainsky, M.A.; Huebner, K.; Croce, C.M.; Vande Woude, G.F. Molecular cloning of a new transforming gene from a chemically transformed human cell line. *Nature* **1984**, *311*, 29–33. [CrossRef] [PubMed]
- 24. Dean, M.; Park, M.; Le Beau, M.M.; Robins, T.S.; Diaz, M.O.; Rowley, J.D.; Blair, D.G.; Vande Woude, G.F. The human met oncogene is related to the tyrosine kinase oncogenes. *Nature* **1985**, *318*, 385–388. [CrossRef] [PubMed]
- 25. Giordano, S.; Ponzetto, C.; Di Renzo, M.F.; Cooper, C.S.; Comoglio, P.M. Tyrosine kinase receptor indistinguishable from the c-met protein. *Nature* **1989**, *339*, 155–156. [CrossRef] [PubMed]
- 26. Birchmeier, C.; Birchmeier, W.; Gherardi, E.; Vande Woude, G.F. Met, metastasis, motility and more. *Nat. Rev. Mol. Cell Biol.* **2003**, *4*, 915–925. [CrossRef] [PubMed]
- 27. Park, M.; Dean, M.; Cooper, C.S.; Schmidt, M.; O'Brien, S.J.; Blair, D.G.; Vande Woude, G.F. Mechanism of met oncogene activation. *Cell* **1986**, 45, 895–904. [CrossRef]
- 28. Nakamura, T.; Nishizawa, T.; Hagiya, M.; Seki, T.; Shimonishi, M.; Sugimura, A.; Tashiro, K.; Shimizu, S. Molecular cloning and expression of human hepatocyte growth factor. *Nature* **1989**, *342*, 440–443. [CrossRef]
- 29. Stoker, M.; Gherardi, E.; Perryman, M.; Gray, J. Scatter factor is a fibroblast-derived modulator of epithelial cell mobility. *Nature* **1987**, 327, 239–242. [CrossRef]
- 30. Bottaro, D.P.; Rubin, J.S.; Faletto, D.L.; Chan, A.M.; Kmiecik, T.E.; Vande Woude, G.F.; Aaronson, S.A. Identification of the hepatocyte growth factor receptor as the c-met proto-oncogene product. *Science* **1991**, 251, 802–804. [CrossRef]
- 31. Gherardi, E.; Chirgadze, D.Y.; Hepple, J.P.; Zhou, H.; Byrd, R.A.; Blundell, T.L. Crystal structure of the NK1 fragment of HGF/SF suggests a novel mode for growth factor dimerization and receptor binding. *Nat. Struct. Biol.* **1999**, *6*, 72–79. [CrossRef]
- 32. Ponzetto, C.; Bardelli, A.; Zhen, Z.; Maina, F.; dalla Zonca, P.; Giordano, S.; Graziani, A.; Panayotou, G.; Comoglio, P.M. A multifunctional docking site mediates signaling and transformation by the hepatocyte growth factor/scatter factor receptor family. *Cell* **1994**, 77, 261–271. [CrossRef]
- 33. Ma, P.C.; Tretiakova, M.S.; Nallasura, V.; Jagadeeswaran, R.; Husain, A.N.; Salgia, R. Downstream signalling and specific inhibition of c-MET/HGF pathway in small cell lung cancer: Implications for tumour invasion. *Br. J. Cancer* **2007**, *97*, 368–377. [CrossRef] [PubMed]
- 34. Zhang, Y.W.; Vande Woude, G.F. HGF/SF-Met signaling in the control of branching morphogenesis and invasion. *J. Cell. Biochem.* **2003**, *88*, 408–417. [CrossRef] [PubMed]
- 35. Weidner, K.M.; Di Cesare, S.; Sachs, M.; Brinkmann, V.; Behrens, J.; Birchmeier, W. Interaction between Gab1 and the c-Met receptor tyrosine kinase is responsible for epithelial morphogenesis. *Nature* **1996**, *384*, 173–176. [CrossRef] [PubMed]
- 36. Bladt, F.; Riethmacher, D.; Isenmann, S.; Aguzzi, A.; Birchmeier, C. Essential role for the c-met receptor in the migration of myogenic precursor cells into the limb bud. *Nature* **1995**, *376*, 768–771. [CrossRef] [PubMed]

Cancers 2019, 11, 1404 10 of 14

37. Huh, C.; Factor, V.M.; Sánchez, A.; Uchida, K.; Conner, E.A.; Thorgeirsson, S.S. Hepatocyte growth factor/c-met signaling pathway is required for efficient liver regeneration and repair. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 4477–4482. [CrossRef]

- 38. Chmielowiec, J.; Borowiak, M.; Morkel, M.; Stradal, T.; Munz, B.; Werner, S.; Wehland, J.; Birchmeier, C.; Birchmeier, W. C-Met is essential for wound healing in the skin. *J. Cell Biol.* **2007**, *177*, 151–162. [CrossRef]
- 39. Peschard, P.; Park, M. Escape from Cbl-mediated downregulation: A recurrent theme for oncogenic deregulation of receptor tyrosine kinases. *Cancer Cell* **2003**, *3*, 519–523. [CrossRef]
- 40. Hammond, D.E.; Urbé, S.; Vande Woude, G.F.; Clague, M.J. Down-regulation of MET, the receptor for hepatocyte growth factor. *Oncogene* **2001**, *20*, 2761–2770. [CrossRef]
- 41. Palka, H.L.; Park, M.; Tonks, N.K. Hepatocyte growth factor receptor tyrosine kinase Met is a substrate of the receptor protein-tyrosine phosphatase DEP-1. *J. Biol. Chem.* **2003**, 278, 5728–5735. [CrossRef]
- 42. Rong, S.; Segal, S.; Anver, M.; Resau, J.H.; Vande Woude, G.F. Invasiveness and metastasis of NIH 3T3 cells induced by Met-hepatocyte growth factor/scatter factor autocrine stimulation. *Proc. Natl. Acad. Sci. USA* **2006**, *91*, 4731–4735. [CrossRef] [PubMed]
- 43. Rong, S.; Bodescot, M.; Blair, D.; Dunn, J.; Nakamura, T.; Mizuno, K.; Park, M.; Chan, A.; Aaronson, S.; Vande Woude, G.F. Tumorigenicity of the met proto-oncogene and the gene for hepatocyte growth factor. *Mol. Cell. Biol.* **2015**, *12*, 5152–5158. [CrossRef] [PubMed]
- 44. Abounader, R.; Lal, B.; Luddy, C.; Koe, G.; Davidson, B.; Rosen, E.M.; Laterra, J. In vivo targeting of SF/HGF and c-met expression via U1snRNA/ribozymes inhibits glioma growth and angiogenesis and promotes apoptosis. *FASEB J.* 2002, *16*, 108–110. [CrossRef] [PubMed]
- 45. Harvey, P.; Warn, A.; Newman, P.; Perry, L.J.; Ball, R.Y.; Warn, R.M. Immunoreactivity for hepatocyte growth factor/scatter factor and its receptor, met, in human lung carcinomas and malignant mesotheliomas. *J. Pathol.* 1996, 180, 389–394. [CrossRef]
- 46. Oh, R.R.; Park, J.Y.; Lee, J.H.; Shin, M.S.; Kim, H.S.; Lee, S.K.; Kim, Y.S.; Lee, S.H.; Lee, S.N.; Yang, Y.M.; et al. Expression of HGF/SF and Met protein is associated with genetic alterations of VHL gene in primary renal cell carcinomas. *APMIS* **2002**, *110*, 229–238. [CrossRef] [PubMed]
- 47. Otte, J.-M.; Schmitz, F.; Kiehne, K.; Stechele, H.U.; Banasiewicz, T.; Krokowicz, P.; Nakamura, T.; Fölsch, U.R.; Herzig, K.-H. Functional Expression of HGF and Its Receptor in Human Colorectal Cancer. *Digestion* **2000**, *61*, 237–246. [CrossRef]
- 48. Edakuni, G.; Sasatomi, E.; Satoh, T.; Tokunaga, O.; Miyazaki, K. Expression of the hepatocyte growth factor/c-Met pathway is increased at the cancer front in breast carcinoma. *Pathol. Int.* **2001**, *51*, 172–178. [CrossRef]
- 49. Cheng, H.-L.; Trink, B.; Tzai, T.-S.; Liu, H.-S.; Chan, S.-H.; Ho, C.-L.; Sidransky, D.; Chow, N.-H. Overexpression of c-met as a Prognostic Indicator for Transitional Cell Carcinoma of the Urinary Bladder: A Comparison with p53 Nuclear Accumulation. *J. Clin. Oncol.* **2002**, *20*, 1544–1550. [CrossRef]
- 50. Joseph, A.; Weiss, G.H.; Jin, L.; Fuchs, A.; Chowdhury, S.; O'Shaugnessy, P.; Goldberg, I.D.; Rosen, E.M. Expression of Scatter Factor in Human Bladder Carcinoma. *JNCI J. Natl. Cancer Inst.* **1995**, *87*, 372–377. [CrossRef]
- 51. Schmidt, L.; Duh, F.M.; Chen, F.; Kishida, T.; Glenn, G.; Choyke, P.; Scherer, S.W.; Zhuang, Z.; Lubensky, I.; Dean, M.; et al. Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. *Nat. Genet.* **1997**, *16*, 68–73. [CrossRef]
- 52. Ma, P.C.; Kijima, T.; Maulik, G.; Fox, E.A.; Sattler, M.; Griffin, J.D.; Johnson, B.E.; Salgia, R. c-MET mutational analysis in small cell lung cancer: Novel juxtamembrane domain mutations regulating cytoskeletal functions. *Cancer Res.* **2003**, *63*, 6272–6281. [PubMed]
- 53. de Aguirre, I.; Salvatierra, A.; Font, A.; Mate, J.L.; Perez, M.; Botia, M.; Taron, M.; Rosell, R. c-Met Mutational Analysis in the Sema and Juxtamembrane Domains in Small-Cell-Lung-Cancer. *Transl. Oncogenom.* **2006**, *1*, 11–18.
- 54. Kong-Beltran, M.; Seshagiri, S.; Zha, J.; Zhu, W.; Bhawe, K.; Mendoza, N.; Holcomb, T.; Pujara, K.; Stinson, J.; Fu, L.; et al. Somatic mutations lead to an oncogenic deletion of Met in lung cancer. *Cancer Res.* **2006**, *66*, 283–289. [CrossRef] [PubMed]
- 55. Tanyi, J.; Tory, K.; Rigó, J.; Nagy, B.; Papp, Z. Evaluation of the tyrosine kinase domain of the Met proto-oncogene in sporadic ovarian carcinomas. *Pathol. Oncol. Res.* **1999**, *5*, 187–191. [CrossRef] [PubMed]

Cancers 2019, 11, 1404 11 of 14

56. Jeffers, M.; Schmidt, L.; Nakaigawa, N.; Webb, C.P.; Weirich, G.; Kishida, T.; Zbar, B.; Vande Woude, G.F. Activating mutations for the Met tyrosine kinase receptor in human cancer. *Proc. Natl. Acad. Sci. USA* **1997**, 94, 11445–11450. [CrossRef] [PubMed]

- 57. Di Renzo, M.F.; Olivero, M.; Martone, T.; Maffe, A.; Maggiora, P.; De Stefani, A.; Valente, G.; Giordano, S.; Cortesina, G.; Comoglio, P.M. Somatic mutations of the MET oncogene are selected during metastatic spread of human HNSC carcinomas. *Oncogene* **2000**, *19*, 1547–1555. [CrossRef] [PubMed]
- 58. Lee, J.-H.; Han, S.-U.; Cho, H.; Jennings, B.; Gerrard, B.; Dean, M.; Schmidt, L.; Zbar, B.; Vande Woude, G.F. A novel germ line juxtamembrane Met mutation in human gastric cancer. *Oncogene* **2000**, *19*, 4947–4953. [CrossRef]
- 59. Craene, B. De Berx, G. Regulatory networks defining EMT during cancer initiation and progression. *Nat. Rev. Cancer* **2013**, *13*, 97–110. [CrossRef]
- 60. Grotegut, S.; Von Schweinitz, D.; Christofori, G.; Lehembre, F. Hepatocyte growth factor induces cell scattering through MAPK/Egr-1-mediated upregulation of Snail. *EMBO J.* **2006**, *25*, 3534–3545. [CrossRef]
- 61. Ogunwobi, O.O.; Liu, C. Hepatocyte growth factor upregulation promotes carcinogenesis and epithelial-mesenchymal transition in hepatocellular carcinoma via Akt and COX-2 pathways. *Clin. Exp. Metastasis* **2011**, *28*, 721–731. [CrossRef]
- 62. Yano, S.; Takeuchi, S.; Nakagawa, T.; Yamada, T. Ligand-triggered resistance to molecular targeted drugs in lung cancer: Roles of hepatocyte growth factor and epidermal growth factor receptor ligands. *Cancer Sci.* **2012**, *103*, 1189–1194. [CrossRef] [PubMed]
- 63. Yano, S.; Wang, W.; Li, Q.; Matsumoto, K.; Sakurama, H.; Nakamura, T.; Ogino, H.; Kakiuchi, S.; Hanibuchi, M.; Nishioka, Y.; et al. Hepatocyte Growth Factor Induces Gefitinib Resistance of Lung Adenocarcinoma with Epidermal Growth Factor Receptor-Activating Mutations. *Cancer Res.* 2008, 68, 9479–9487. [CrossRef] [PubMed]
- 64. Finisguerra, V.; Prenen, H.; Mazzone, M. Preclinical and clinical evaluation of MET functions in cancer cells and in the tumor stroma. *Oncogene* **2016**, *35*, 5457–5467. [CrossRef] [PubMed]
- 65. Gherardi, E.; Birchmeier, W.; Birchmeier, C.; Woude, G. Vande Targeting MET in cancer: Rationale and progress. *Nat. Rev. Cancer* **2012**, *12*, 89–103. [CrossRef] [PubMed]
- 66. Zhang, Y.-W.; Su, Y.; Volpert, O.V.; Woude, G.F.V. Hepatocyte growth factor/scatter factor mediates angiogenesis through positive VEGF and negative thrombospondin 1 regulation. *Proc. Natl. Acad. Sci. USA* 2003, 100, 12718–12723. [CrossRef] [PubMed]
- 67. Bharti, A.; Ma, P.C.; Maulik, G.; Singh, R.; Khan, E.; Skarin, A.T.; Salgia, R. Haptoglobin α-Subunit and Hepatocyte Growth Factor can Potentially Serve as Serum Tumor Biomarkers in Small Cell Lung Cancer. *Anticancer Res.* **2004**, 24, 1031–1038. [PubMed]
- 68. Masuya, D.; Huang, C.; Liu, D.; Nakashima, T.; Kameyama, K.; Haba, R.; Ueno, M.; Yokomise, H. The tumour-stromal interaction between intratumoral c-Met and stromal hepatocyte growth factor associated with tumour growth and prognosis in non-small-cell lung cancer patients. *Br. J. Cancer* **2004**, *90*, 1555–1562. [CrossRef]
- 69. Ghoussoub, R.A.D.; Dillon, D.A.; D'Aquila, T.; Rimm, E.B.; Fearon, E.R.; Rimm, D.L. Expression of c-met is a strong independent prognostic factor in breast carcinoma. *Cancer* **1998**, *82*, 1513–1520. [CrossRef]
- 70. Cañadas, I.; Taus, A.; González, I.; Villanueva, X.; Gimeno, J.; Pijuan, L.; Dómine, M.; Sánchez-Font, A.; Vollmer, I.; Menéndez, S.; et al. High circulating hepatocyte growth factor levels associate with epithelial to mesenchymal transition and poor outcome in small cell lung cancer patients. *Oncotarget* 2014, 5, 5246–5256. [CrossRef]
- 71. Rygaard, K.; Nakamura, T.; Spang-Thomsen, M. Expression of the proto-oncogenes c-met and c-kit and their ligands, hepatocyte growth factor/scatter factor and stem cell factor, in sclc cell lines and xenografts. *Br. J. Cancer* 1993, 67, 37–46. [CrossRef]
- 72. Maulik, G.; Kijima, T.; Ma, P.C.; Ghosh, S.K.; Lin, J.; Shapiro, G.I.; Schaefer, E.; Tibaldi, E.; Johnson, B.E.; Salgia, R. Modulation of the c-Met/hepatocyte growth factor pathway in small cell lung cancer. *Clin. Cancer Res.* **2002**, *8*, 620–627. [PubMed]
- 73. Webb, C.P.; Hose, C.D.; Koochekpour, S.; Jeffers, M.; Oskarsson, M.; Sausville, E.; Monks, A.; Vande Woude, G.F. The geldanamycins are potent inhibitors of the hepatocyte growth factor/scatter factor-Met-urokinase plasminogen activator-plasmin proteolytic network. *Cancer Res.* **2000**, *60*, 342–349. [PubMed]

Cancers 2019, 11, 1404 12 of 14

74. Cho, S.H.; Kim, J.I.; Kim, H.S.; Park, S.D.; Jang, K.W. The antitumor effect of c-terminus of hsp70-interacting protein via degradation of c-met in small cell lung cancer. *Korean J. Thorac. Cardiovasc. Surg.* **2017**, *50*, 153–162. [CrossRef] [PubMed]

- 75. Maulik, G.; Madhiwala, P.; Brooks, S.; Ma, P.C.; Kijima, T.; Tibaldi, E.V.; Schaefer, E.; Parmar, K.; Salgia, R. Activated c-Met signals through PI3K with dramatic effects on cytoskeletal functions in small cell lung cancer. *J. Cell. Mol. Med.* **2002**, *6*, 539–553. [CrossRef] [PubMed]
- 76. Peruzzi, B.; Bottaro, D.P. Targeting the c-Met signaling pathway in cancer. *Clin. Cancer Res.* **2006**, 12, 3657–3660. [CrossRef] [PubMed]
- 77. Sattler, M.; Pride, Y.B.; Ma, P.; Gramlich, J.L.; Chu, S.C.; Quinnan, L.A.; Shirazian, S.; Liang, C.; Podar, K.; Christensen, J.G.; et al. A novel small molecule Met inhibitor induces apoptosis in cells transformed by the oncogenic TPR-MET tyrosine kinase. *Cancer Res.* **2003**, *63*, 5462–5469.
- 78. Ma, P.C.; Jagadeeswaran, R.; Jagadeesh, S.; Tretiakova, M.S.; Nallasura, V.; Fox, E.A.; Hansen, M.; Schaefer, E.; Naoki, K.; Lader, A.; et al. Functional expression and mutations of c-Met and its therapeutic inhibition with SU11274 and small interfering RNA in non-small cell lung cancer. *Cancer Res.* **2005**, *65*, 1479–1488. [CrossRef] [PubMed]
- 79. Ma, P.C.; Schaefer, E.; Christensen, J.G.; Salgia, R. A selective small molecule c-MET inhibitor, PHA665752, cooperates with rapamycin. *Clin. Cancer Res.* **2005**, *11*, 2312–2319. [CrossRef] [PubMed]
- 80. Puri, N.; Khramtsov, A.; Ahmed, S.; Nallasura, V.; Hetzel, J.T.; Jagadeeswaran, R.; Karczmar, G.; Salgia, R. A selective small molecule inhibitor of c-Met, PHA665752, inhibits tumorigenicity and angiogenesis in mouse lung cancer xenografts. *Cancer Res.* **2007**, *67*, 3529–3534. [CrossRef] [PubMed]
- 81. Rolle, C.E.; Kanteti, R.; Surati, M.; Nandi, S.; Dhanasingh, I.; Yala, S.; Tretiakova, M.; Arif, Q.; Hembrough, T.; Brand, T.M.; et al. Combined MET Inhibition and Topoisomerase I Inhibition Block Cell Growth of Small Cell Lung Cancer. *Mol. Cancer Ther.* **2013**, *13*, 576–584. [CrossRef]
- 82. Kawato, Y.; Aonuma, M.; Hirota, Y.; Kuga, H.; Sato, K. Intracellular roles of SN-38, a metabolite of the camptothecin derivative CPT-11, in the antitumor effect of CPT-11. *Cancer Res.* **1991**, *51*, 4187–4191. [PubMed]
- 83. Ettinger, D.S. New drugs for treating small cell lung cancer. Lung Cancer 1995, 12, S53–S61. [CrossRef]
- 84. Christensen, J.G.; Zou, H.Y.; Arango, M.E.; Li, Q.; Lee, J.H.; McDonnell, S.R.; Yamazaki, S.; Alton, G.R.; Mroczkowski, B.; Los, G. Cytoreductive antitumor activity of PF-2341066, a novel inhibitor of anaplastic lymphoma kinase and c-Met, in experimental models of anaplastic large-cell lymphoma. *Mol. Cancer Ther.* **2007**, *6*, 3314–3322. [CrossRef] [PubMed]
- 85. Ozasa, H.; Oguri, T.; Maeno, K.; Takakuwa, O.; Kunii, E.; Yagi, Y.; Uemura, T.; Kasai, D.; Miyazaki, M.; Niimi, A. Significance of c-MET overexpression in cytotoxic anticancer drug-resistant small-cell lung cancer cells. *Cancer Sci.* **2014**, *105*, 1032–1039. [CrossRef] [PubMed]
- 86. Sakamoto, S.; Inoue, H.; Ohba, S.; Kohda, Y.; Usami, I.; Masuda, T.; Kawada, M.; Nomoto, A. New metastatic model of human small-cell lung cancer by orthotopic transplantation in mice. *Cancer Sci.* **2015**, *106*, 367–374. [CrossRef] [PubMed]
- 87. Taniguchi, H.; Yamada, T.; Takeuchi, S.; Arai, S.; Fukuda, K.; Sakamoto, S.; Kawada, M.; Yamaguchi, H.; Mukae, H.; Yano, S. Impact of MET inhibition on small-cell lung cancer cells showing aberrant activation of the hepatocyte growth factor/MET pathway. *Cancer Sci.* **2017**, *108*, 1378–1385. [CrossRef] [PubMed]
- 88. Nakagawa, T.; Tohyama, O.; Yamaguchi, A.; Matsushima, T.; Takahashi, K.; Funasaka, S.; Shirotori, S.; Asada, M.; Obaishi, H. E7050: A dual c-Met and VEGFR-2 tyrosine kinase inhibitor promotes tumor regression and prolongs survival in mouse xenograft models. *Cancer Sci.* **2010**, *101*, 210–215. [CrossRef] [PubMed]
- 89. Basilico, C.; Pennacchietti, S.; Vigna, E.; Chiriaco, C.; Arena, S.; Bardelli, A.; Valdembri, D.; Serini, G.; Michieli, P. Tivantinib (ARQ197) displays cytotoxic activity that is independent of its ability to bind MET. *Clin. Cancer Res.* **2013**, *19*, 2381–2392. [CrossRef] [PubMed]
- 90. Calles, A.; Kwiatkowski, N.; Cammarata, B.K.; Ercan, D.; Gray, N.S.; Jänne, P.A. Tivantinib (ARQ 197) efficacy is independent of MET inhibition in non-small-cell lung cancer cell lines. *Mol. Oncol.* **2015**, *9*, 260–269. [CrossRef]
- 91. Liu, S.V.; Groshen, S.G.; Kelly, K.; Reckamp, K.L.; Belani, C.; Synold, T.W.; Goldkorn, A.; Gitlitz, B.J.; Cristea, M.C.; Gong, I.Y.; et al. A phase I trial of topotecan plus tivantinib in patients with advanced solid tumors. *Cancer Chemother. Pharmacol.* **2018**, *82*, 723–732. [CrossRef]

Cancers 2019, 11, 1404 13 of 14

92. US National Library of Medicine. ClinicalTrials.gov. 2017. Available online: https://clinicaltrials.gov/ct2/show/NCT02608411 (accessed on 11 May 2019).

- 93. Glisson, B.; Besse, B.; Dols, M.C.; Dubey, S.; Schupp, M.; Jain, R.; Jiang, Y.; Menon, H.; Nackaerts, K.; Orlov, S.; et al. A Randomized, Placebo-Controlled, Phase 1b/2 Study of Rilotumumab or Ganitumab in Combination With Platinum-Based Chemotherapy as First-Line Treatment for Extensive-Stage Small-Cell Lung Cancer. *Clin. Lung Cancer* 2017, 18, 615–625.e8. [CrossRef] [PubMed]
- 94. Byers, L.A.; Horn, L.; Ghandi, J.; Kloecker, G.; Owonikoko, T.; Waqar, S.N.; Krzakowski, M.; Cardnell, R.J.; Fujimoto, J.; Taverna, P.; et al. A phase 2, open-label, multi-center study of amuvatinib in combination with platinum etoposide chemotherapy in platinum-refractory small cell lung cancer patients. *Oncotarget* **2017**, *8*, 81441–81454. [CrossRef] [PubMed]
- 95. Peifer, M.; Fernández-Cuesta, L.; Sos, M.L.; George, J.; Seidel, D.; Kasper, L.H.; Plenker, D.; Leenders, F.; Sun, R.; Zander, T.; et al. Integrative genome analyses identify key somatic driver mutations of small-cell lung cancer. *Nat. Genet.* **2012**, *44*, 1104–1110. [CrossRef] [PubMed]
- 96. Bordi, P.; Tiseo, M.; Barbieri, F.; Bavieri, M.; Sartori, G.; Marchetti, A.; Buttitta, F.; Bortesi, B.; Ambrosini-Spaltro, A.; Gnetti, L.; et al. Gene mutations in small-cell lung cancer (SCLC): Results of a panel of 6 genes in a cohort of Italian patients. *Lung Cancer* 2014, 86, 324–328. [CrossRef] [PubMed]
- 97. Voortman, J.; Harada, T.; Chang, R.P.; Killian, J.K.; Suuriniemi, M.; Smith, W.I.; Meltzer, P.S.; Lucchi, M.; Wang, Y.; Giaccone, G. Detection and Therapeutic Implications of c-Met Mutations in Small Cell Lung Cancer and Neuroendocrine Tumors. *Curr. Pharm. Des.* 2013, 19, 833–840. [CrossRef] [PubMed]
- 98. Sakai, K.; Passioura, T.; Sato, H.; Ito, K.; Furuhashi, H.; Umitsu, M.; Takagi, J.; Kato, Y.; Mukai, H.; Warashina, S.; et al. Macrocyclic peptide-based inhibition and imaging of hepatocyte growth factor. *Nat. Chem. Biol.* **2019**, *15*, 598–606. [CrossRef] [PubMed]
- 99. Schag, K.; Schmidt, S.M.; Müller, M.R.; Weinschenk, T.; Appel, S.; Weck, M.M.; Grünebach, F.; Stevanovic, S.; Rammensee, H.G.; Brossart, P. Identification of C-Met oncogene as a broadly expressed tumor-associated antigen recognized by cytotoxic T-lymphocytes. *Clin. Cancer Res.* **2004**, *10*, 3658–3666. [CrossRef]
- 100. Liu, Y.; Wilkinson, F.L.; Kirton, J.P.; Jeziorska, M.; Iizasa, H.; Sai, Y.; Nakashima, E.; Heagerty, A.M.; Canfield, A.E.; Alexander, M.Y. Hepatocyte growth factor and c-Met expression in pericytes: Implications for atherosclerotic plaque development. *J. Pathol.* 2007, 212, 12–19. [CrossRef]
- 101. Chen, Q.; De, F.M.; Zarnegar, R. Induction of met proto-oncogene (hepatocyte growth factor receptor) expression during human monocyte-macrophage differentiation. *Cell Growth Differ* **1996**, 7, 821–832.
- 102. Benkhoucha, M.; Santiago-Raber, M.-L.; Schneiter, G.; Chofflon, M.; Funakoshi, H.; Nakamura, T.; Lalive, P.H. Hepatocyte growth factor inhibits CNS autoimmunity by inducing tolerogenic dendritic cells and CD25+Foxp3+ regulatory T cells. *Proc. Natl. Acad. Sci. USA* 2010, 107, 6424–6429. [CrossRef]
- 103. Benkhoucha, M.; Molnarfi, N.; Kaya, G.; Belnoue, E.; Bjarnadóttir, K.; Dietrich, P.; Walker, P.R.; Martinvalet, D.; Derouazi, M.; Lalive, P.H. Identification of a novel population of highly cytotoxic c-Met-expressing CD8 + T lymphocytes. *EMBO Rep.* **2017**, *18*, 1545–1558. [CrossRef]
- 104. Finisguerra, V.; Di Conza, G.; Di Matteo, M.; Serneels, J.; Costa, S.; Thompson, A.A.R.; Wauters, E.; Walmsley, S.; Prenen, H.; Granot, Z.; et al. MET is required for the recruitment of anti-tumoural neutrophils. *Nature* **2015**, *522*, 349–353. [CrossRef]
- 105. Coffelt, S.B.; Kersten, K.; Doornebal, C.W.; Weiden, J.; Vrijland, K.; Hau, C.S.; Verstegen, N.J.M.; Ciampricotti, M.; Hawinkels, L.J.A.C.; Jonkers, J.; et al. IL-17-producing γδ T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* **2015**, *5*22, 345–348. [CrossRef]
- 106. Glodde, N.; Bald, T.; van den Boorn-Konijnenberg, D.; Nakamura, K.; O'Donnell, J.S.; Szczepanski, S.; Brandes, M.; Eickhoff, S.; Das, I.; Shridhar, N.; et al. Reactive Neutrophil Responses Dependent on the Receptor Tyrosine Kinase c-MET Limit Cancer Immunotherapy. *Immunity* **2017**, 47, 789–802.e9. [CrossRef]
- 107. Schrock, A.B.; Li, S.D.; Frampton, G.M.; Suh, J.; Braun, E.; Mehra, R.; Buck, S.C.; Bufill, J.A.; Peled, N.; Karim, N.A.; et al. Pulmonary Sarcomatoid Carcinomas Commonly Harbor Either Potentially Targetable Genomic Alterations or High Tumor Mutational Burden as Observed by Comprehensive Genomic Profiling. *J. Thorac. Oncol.* 2017, 12, 932–942. [CrossRef]
- 108. Sabari, J.K.; Leonardi, G.C.; Shu, C.A.; Umeton, R.; Montecalvo, J.; Ni, A.; Chen, R.; Dienstag, J.; Mrad, C.; Bergagnini, I.; et al. PD-L1 expression, tumor mutational burden, and response to immunotherapy in patients with MET exon 14 altered lung cancers. *Ann. Oncol.* **2018**, *29*, 2085–2091. [CrossRef]

Cancers **2019**, 11, 1404

109. Greenall, S.A.; Adams, T.E.; Johns, T.G. Incomplete target neutralization by the anti-cancer antibody rilotumumab. *MAbs* **2016**, *8*, 246–252. [CrossRef]

110. Adjei, A.A.; Schwartz, B.; Garmey, E. Early Clinical Development of ARQ 197, a Selective, Non-ATP-Competitive Inhibitor Targeting MET Tyrosine Kinase for the Treatment of Advanced Cancers. *Oncologist* 2011, 16, 788–799. [CrossRef]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).